

RESEARCH

Open Access

A Thymidine Kinase recombinant protein-based ELISA for detecting antibodies to Duck Plague Virus

Yongping Wen¹, Anchun Cheng^{*1,2,3}, Mingshu Wang^{*1,2}, Han Ge¹, Chanjuan Shen¹, Sitong Liu¹, Jun Xiang¹, Renyong Jia², Dekang Zhu², Xiaoyue Chen^{1,3}, Bei Lian¹, Hua Chang¹ and Yi Zhou²

Abstract

Background: Duck plague virus (DPV) is the causative agent of Duck Plague (DP) that causes significant morbidity and mortality throughout duck-producing areas of the world. The diagnosis of DP currently relies on the use of live or inactivated whole DPV virion as antigens in ELISA, but it is too laborious and expensive for routine application, and it is still difficult to get purified DPV virion with current technology.

Results: In this study, we describe the expression and purification of a recombinant Thymidine Kinase (TK) protein which makes antigen in an in-house developed, optimized and standardized ELISA. The specificity of the optimized TK-ELISA was evaluated by antisera against Duck Plague Virus (DPV), Duck Hepatitis B Virus (DHBV), Duck Hepatitis Virus (DHV), *Riemerella Anatipestifer* (R. A), *Escherichia coli* (E. coli) and *Salmonella anatum* (S. anatum). Only antisera against DPV yielded a specific and strong signal. In order to determine the sensitivity of the TK-ELISA, a panel of diluted sera was tested, and the minimum detection limit of 1:2560 (OD_{450 nm} = 0.401) was obtained according to the endpoint cut-off (0.2438). The repeatability and reproducibility under the experimental conditions demonstrates a low variability ($P > 0.05$). The suspected sera samples ($n = 30$) were determined by TK-ELISA and the positive rate is 90% (27/30), and the TK-ELISA showed 83.33% (22+3/30) coincidence rate with the Serum Neutralization Test (SNT) and 90% (24+3/30) coincidence rate with the whole DPV virion based-ELISA (DPV-ELISA). When defining the dynamics of antibody response to attenuated live DPV vaccine, the maximum antibodies is reached after 4 weeks.

Conclusions: The results suggest that the TK-ELISA provides high specificity, sensitivity, repeatability and reproducibility for detection of anti-DPV antibodies in duck sera, and has the potential to be much simpler than DPV-ELISA and SNT for the sera epidemiological investigation.

Background

Duck plague (DP), which is caused by Duck Plague Virus (DPV), is an acute, contagious and lethal disease discovered, firstly, in Holland [1]. DPV is currently classified belonging to the Alphaherpesvirinae subfamily of the herpesvirus family but has not been grouped into any genus yet [2]. There are 34 species within Order Anseriformes' host range. Ducks, geese, and swans are the susceptible species to DP [3]. The characteristics of DP are vascular damage, tissue hemorrhages, digestive mucosal

eruptions, lesions of lymphoid organs, and degenerative changes in parenchymatous organ [4,5]. Considerable economic losses were suffered by the DP in duck-producing areas of the world [6,7]. China, holding the largest population of waterfowl [8], was also inflicted with heavy losses attributing to the outbreak of DP, since it was firstly reported by Huang [9]. Therefore, to develop a fast and available diagnose method to predict the infection of DPV in a suspected flock, on farm, is extremely urgent.

The diagnosis of DP may be made on the basis of characteristic internal lesions and final diagnosis can be made by virus isolation and identification [10,11], however, it is laborious and time-consuming. In recent years, the Fluorescent Quantitative Real-Time PCR (FQ-PCR) [12],

* Correspondence: chengananchun@vip.163.com, mswang@163.com

¹ Avian Diseases Research Center, College of Veterinary Medicine of Sichuan Agricultural University, Ya'an, Sichuan, 625014, China

[†] Contributed equally

Full list of author information is available at the end of the article

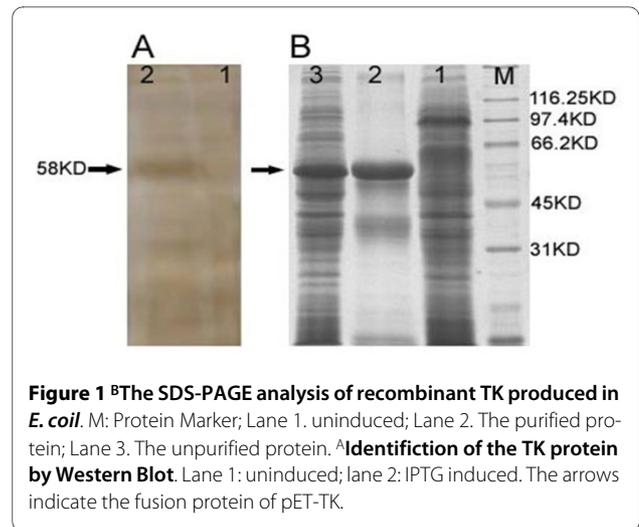
Loop-Mediated Isothermal Amplification(LAMP) [13], Antigen-Capture Enzyme-Linked Immunosorbent Assay (AC-ELISA) [14], Histopathology [15] and Electron Microscopy [16] have been developed quickly. Whereas the key of prevention and control is more than clinical diagnosis, the vaccination, also a critical factor, is generally considered to be the most effective and financially viable method of preventing infectious diseases. In vaccination against DP, antibody detection plays an extremely important role. It is used to detect (subclinical) field infections, to check the response to vaccines and predict the optimal age for vaccination. The gold standard assay for DP antibody detection is the Neutralization Test (NT) [17]. However, the NT requires maintenance and use of live virus and cell cultures and must be performed under aseptic conditions, furthermore, it requires up to 3 days for results. Until now the ELISA assays have been developed for antibody or antigen detection [18-20] and the whole DPV virion usually acts as antigen for the detection of antibodies against DPV in the indirect ELISA assay (I-ELISA) [21]. But much time and energy must be paid in getting the virion as the coating antigen. Compared with the DPV-ELISA, the development of the TK-ELISA assay in this paper is more economical and more convenient.

The DPV gene library has been constructed and identified [22]. The Thymidine Kinase (TK) gene of DPV has been successfully cloned [23]. Comparison with other available Herpesvirus, TK sequences reveals a high homology to those of the Alphaherpesviruses [23-26]. The research also shows that the TK gene, considered the main virulence gene [27,28], is dispensable during the process of virus multiplication [29]. Therefore, the aim of this paper was to describe the expression of a recombinant TK protein, which is applied in antigen coating, in an in-house developed, optimized and standardized ELISA. To date, there are no reports of using expressed DPV recombinant protein acting the same role. To the best of our knowledge, this is the first report of expressing TK protein of DPV in *Escherichia coli* (*E. coli*) and the development of an indirect ELISA test.

Results

Expression of the recombinant DPV TK protein

The product was expressed in *E. coli* system as a His6-tagged recombinant TK fusion protein of approximately 58 KD (Fig. 1.B, lane 2,3). Western Blot analysis showed that the purified His6-tagged TK was recognized by the rabbit anti-DPV IgG with a specific signal at 58 KD; which is the expected size of the fusion protein (Fig. 1.A, lane 2). However, when using the negative control serum, no positive signal was observed (Fig. 1.A, lane 1); indicating that the recombinant protein induced an immunolog-



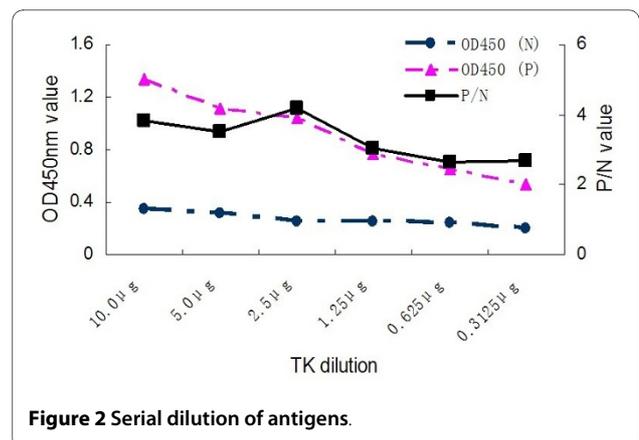
ical response and thus the antiserum had a high level of specificity.

Optimization of ELISA Procedure

The coating antigen and serum dilution corresponding to the highest P/N value (ODtest sample/ODnegative control) were regarded as optimal antigen concentration and optimal serum dilution. The optimal antigen concentration of TK protein, anti-duck horseradish peroxidase secondary antibody dilution and the serum dilution were found by checkerboard titration at 2.5 $\mu\text{g}/100 \mu\text{l}$ (1:200) (Fig. 2), 1:2000 and 1:80 (Fig. 4), respectively. The process of optimization of the anti-duck horseradish peroxidase secondary antibody and sera dilutions was revealed in the Fig. 3, 4 and 5.

The endpoint cut-off value

The results of intra-day and inter-day assays analyzed by SAS8.22 showed the difference among 6 groups was not significant ($P > 0.05$). The OD_{450 nm} average values of 0.1211 was acquired after these duck-negative sera samples ($n = 30$) were detected by the TK-ELISA with a stan-



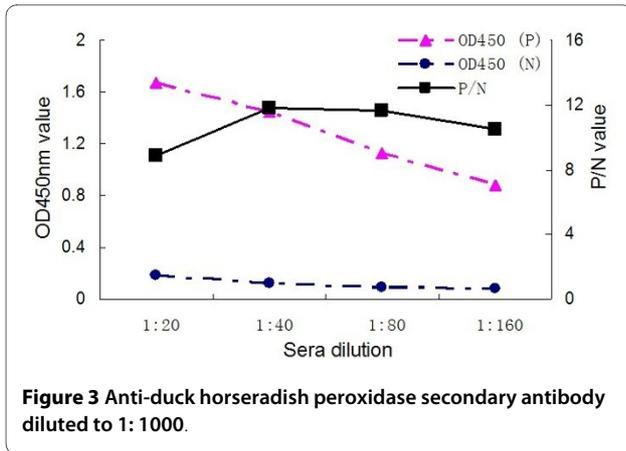


Figure 3 Anti-duck horseradish peroxidase secondary antibody diluted to 1: 1000.

standard deviation of 0.0409. The endpoint cut-off value of the ELISA was calculated according to a Gaussian population distribution [30]. Consequently, for a 99% confidence interval, the endpoint cut-off value was defined as follows: mean of the negative serum OD_{450 nm} values plus three standard deviations = 0.1211 + 3 × 0.0409 = 0.2438.

Analytical specificity and sensitivity of the TK-ELISA

On the basis of endpoint cut-off value (0.2438), the OD_{450 nm} value of the DPV-positive sera is 1.398 and 1.432 both of which were higher than 0.2438, therefore, positive. The OD_{450 nm} value of the other samples were all lower than 0.2438. The results indicated that TK-ELISA had a high specificity for the DPV-positive sera and there was no cross-reaction with antisera against Duck Hepatitis B Virus (DHBV), Duck Hepatitis Virus (DHV), *Riemerella Anatipestifer* (*R. A*), *Escherichia coli* (*E. coli*) and *Salmonella anatum* (*S. anatum*) (Fig. 6).

After evaluating the sensitivity of a panel of diluted sera, a minimum detection limit of 1:2560 (OD_{450 nm} = 0.401) was obtained according to the endpoint cut-off (0.2438), but the blank control did not yield positive results (Table 1).

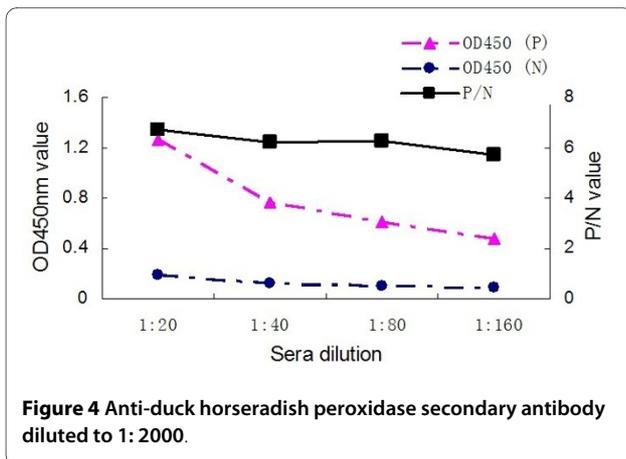


Figure 4 Anti-duck horseradish peroxidase secondary antibody diluted to 1: 2000.

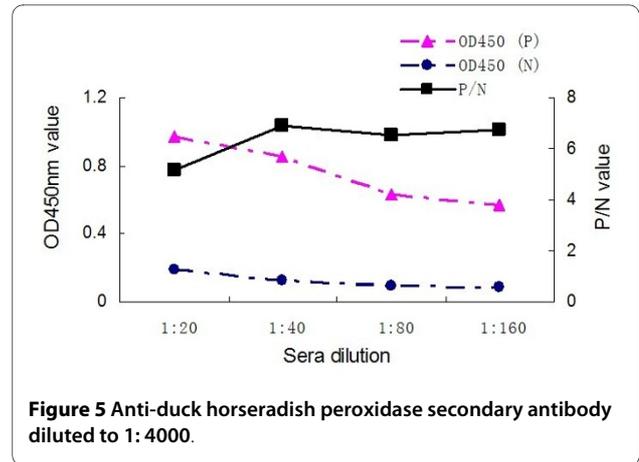


Figure 5 Anti-duck horseradish peroxidase secondary antibody diluted to 1: 4000.

Repeatability and reproducibility of the TK-ELISA

The results analyzed by SAS8.22 showed the difference among 3 groups was not significant ($P > 0.05$). Table 2, 3 shows the repeatability and reproducibility of the assay under the experimental conditions, demonstrating the low variability.

Analytical inhibition of the TK-ELISA

After repeating three times using the TK-ELISA, it was observed that the OD_{450 nm} values in these reactions declined sharply by more than 50% after blocking with TK protein (Fig. 7). This result indicates that the assay was highly specific for DPV-TK antigen.

The coincidence rate among SNT, TK-ELISA and DPV-ELISA

By the TK-ELISA, the SNT and DPV-ELISA, the suspected sera samples ($n = 30$) were determined, respectively. The results showed that the positive rate were 90% (27/30), 80% (24/30) and 73.3% (22/30), respectively. In addition, the TK-ELISA showed 83.33% (22+3/30) coincidence rate with the SNT and 90% (24+3/30) coincidence rate with the DPV-ELISA (Table 4).

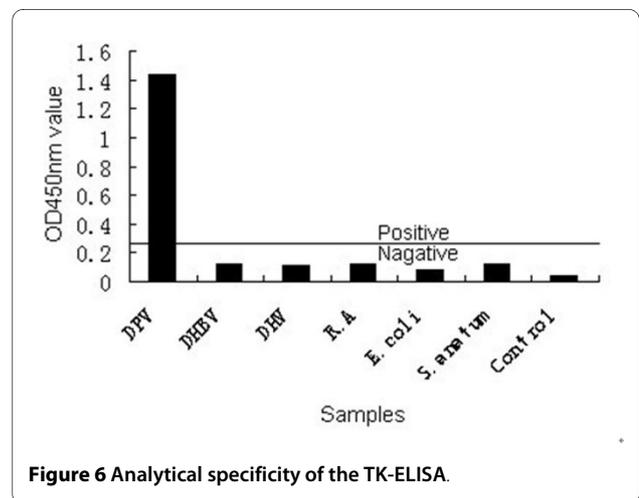


Figure 6 Analytical specificity of the TK-ELISA.

Table 1: One positive serum sample with gradient dilution to evaluate the sensitivity of the TK-ELISA.

Dilution	1:160	1:320	1:640	1:1280	1:2560	1:5120	1:10240
OD _{450 nm}	1.432	0.984	0.975	0.542	0.401	0.107	0.111
P or N	+	+	+	+	+	-	-

The sera-positive and sera-negative are marked with a plus (+) or a minus (-).

The kinetics of antibody production

The dynamics of antibody response to attenuated live DPV vaccine for duck using the TK-ELISA and the DPV-ELISA are shown in Fig 8. In terms of TK-ELISA, the OD_{450 nm} values started to rise at approximately day 5, and after inoculation reaching maximum value at week 4, followed, after, by their gradual decrease. But as for DPV-ELISA, the OD_{450 nm} values started to rise at approximately day 7, which is a bit later than that of TK-ELISA.

Discussion

In this study, the TK recombinant protein was produced in *E. coli* system to replace the whole DPV virion used for detecting antibodies in TK-ELISA. The use of un-purified whole virion preparation in the I-ELISA, brings high background absorbance but purification of the virion is laborious and expensive [31]. Therefore, many people choose the recombinant protein which has advantages over whole virion as antigen for the detection of antibodies [32-35]. Firstly, the recombinant protein antigen is not infectious, so it gives the opportunity to guarantee DPV serodiagnostic assays rapidly and safely without virus containment facilities. Secondly, the recombinant protein antigen could be easily produced in general laboratory without any limitation. Particularly, with the help of recombinant antigen coated on the plates, specificity of the ELISA was greatly improved. This enhanced specificity is especially important for eradication programs in developing countries where true-positive results are often interfered due to the deficient conditions of sera transport and storage; one of the most common reasons for unspecific results in ELISA [36].

The TK gene, which is the main virulence gene and dispensable gene for herpesvirus multiplication, is essential

for latent infection [37,38]. Consequently, the TK gene is often considered to be the prior target gene for constructing the gene-deleted vaccine. Based on the deletion of TK gene, the vaccine of Pseudorabies Virus (PRV) [39] and Bovine Herpesvirus (BHV) [40] have been developed successfully. It suggests that preventing and controlling DPV may depend on the TK gene-deleted vaccine in future. Previously, the related reports present that the protein-based ELISA can be used to distinguish antibodies induced by the wild-type virus strain infection from those induced by vaccinated animals [41-43]. Therefore, if DPV TK gene-deleted vaccine was developed successfully, the TK-ELISA discussed above would significantly contribute to the diagnosis methods of discrimination between the infection of DPV TK gene-deleted vaccine strain and that of wild-type DPV strain. Similar to the PRV eradication programs, it can strongly support that of DPV in future.

Another, the *E. coli* system, was chosen to produce the TK protein. The many advantages of *E. coli* have ensured that it remains a valuable organism for the high-level production of recombinant proteins [44,45]. The TK protein, which is highly conserved among DPV can be used as candidate diagnostic antigen because of its group and type specificity. Purification of expressed TK protein by Nickel Affinity Chromatography is able to insure the unique component of coating protein, as well as the decrease of non-specific binding. Furthermore, the purified protein was gradient dialysed to reduce concentration of urea which is added at the time of dissolving inclusion bodies [46]. Thus, the renaturation of recombinant protein makes good antigenicity for coating antigen. To enhance the specificity and sensitivity of the TK-ELISA, the dilution of the TK protein and anti-duck horseradish peroxidase secondary antibody were

Table 2: The repeatability of the assay.

NO.	Number of replications						X	SD	C.V(%)
	1	2	3	4	5	6			
A	1.322	1.485	1.409	1.388	1.425	1.605	1.439	0.097	6.742
B	1.611	1.569	1.398	1.487	1.552	1.623	1.540	0.085	5.501
C	1.324	1.382	1.578	1.418	1.308	1.509	1.419	0.106	7.459

Table 3: The reproducibility of the assay.

Days.	NO.			X	SD	C.V(%)
	A	B	C			
1	1.401	1.609	1.535	1.515	0.105	6.959
2	1.602	1.498	1.487	1.529	0.063	4.150
3	1.384	1.332	1.517	1.411	0.095	6.762

screened and the optimal dilutions are 2.5 µg/100 µl (1:200) and 1:2 000, respectively. What's more, Steinitz et al. [47] presents a practical approach to defining the conditions on optimal coating, such as coating buffer, temperature and duration of coating. However, although there is much research into the effects of temperature and duration of coating, there is little research into the effects of coating buffer. In this study, in order to get the optimal coating buffer, 0.05 M bicarbonate/carbonate buffer (pH 9.6), Phosphate Buffered Saline (pH 7.4 and 8.5) were screened. It was discovered that 0.05 M bicarbonate/carbonate buffer (pH 9.6) was the best coating buffer for immobilization of the protein. Otherwise, we have found that coating plates at 4°C overnight also reduced the background.

Interestingly as an additional discovery, the DPV-specific antibodies could be clearly detected approximately 5 days earlier than the conventional assay after attenuated live DPV vaccine-induced immunity. Most likely, TK gene is a part of early genes of herpesvirus, in which TK genes share the high conservation [48,49]. The TK, as the key enzyme in replication of DPV, is probably expressed at early times after infection, which caused its rapid immune recognition after the infection.

In summary; in this study, the TK-ELISA manages to provide an new approach to screening and studying DPV, and this method could replace DPV-ELISA and SNT as an alternative, sometimes prove even more effective.

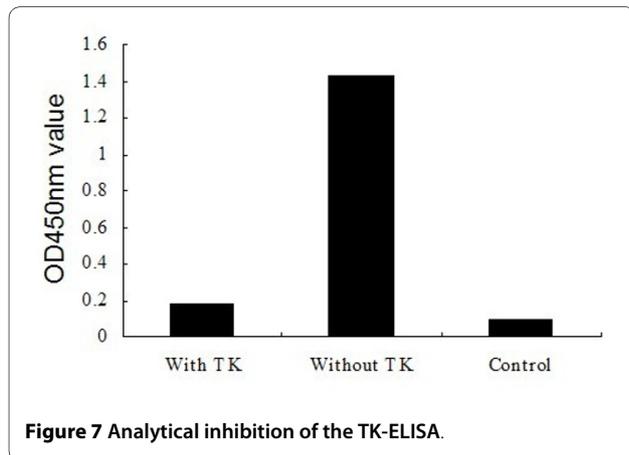


Figure 7 Analytical inhibition of the TK-ELISA.

Conclusions

This paper demonstrated the expression of the TK protein of DPV in *E. coli* system and their application in developing an indirect TK-ELISA to detect anti-DPV antibodies from DPV-infected duck sera. The TK-ELISA, as a simple and economical method, may exert a strong influence on DPV eradication campaigns and control programs in duck-producing areas of the world.

Methods

Sera samples

A panel of sera containing 70 duck sera samples, of which 5 were DPV-positive, 35 were DPV-negative, and remaining 30 were suspected infectious DPV, was used in this study. The 30 suspected DPV-positive sera samples were obtained from clinical samples. The 5 DPV-positive sera were acquired by inoculation of DPV Virulence Strain. 30 of the sera samples from ducks uninfected with DPV (DPV-negative) were used to evaluate the endpoint cut-off value of TK-ELISA, and the remaining 5 DPV-negative serum were used to optimize the procedure of TK-ELISA. To calculate the specificity of the TK-ELISA 20 serum samples tested positive for DHBV, DHV, *R. A*, *E. coli*, and infectious *S. anatum* were also used. To study the kinetics of antibody production 20 serum samples were obtained by inoculating a new different group of DPV infected ducks subcutaneously with the attenuated live DPV vaccine. All the test sera above were collected and were preserved at -70°C in author's laboratory until used.

Expression and purification of recombinant DPV TK protein

The pET-32a(+)/Tk expression plasmid was provided by the author's laboratory [23]. The pET-32a(+)-Tk was transformed into *E. coli* BL21(DE3) and the recombinant clones were selected on Luria bertani (LB) agar plates containing ampicillin (100 mg/ml). Transformants grew on LB plates containing ampicillin (100 µg/ml) at 37°C for 18 h. A single colony from the culture was inoculated into LB medium with ampicillin. When the cultures initially reached logarithmic phase (at OD₆₀₀ of 0.5-0.6), protein expression was induced at 37°C by isopropyl-β-D-thiogalactoside (IPTG) (final concentration 0.6 mM) for a fur-

Table 4: The coincidence rate among SNT, TK-ELISA and DPV-ELISA for detection of antibodies to DPV.

TK-ELISA	SNT		DPV-ELISA		Total
	Positive	Negative	Positive	Negative	
Positive	22	5	24	3	27
Negative	0	3	0	3	3
Total	22	8	24	6	30

The coincidence rate was calculated by the formula: (no. of samples positive by both method + no. of samples negative by both methods)/total no. samples× 100.

ther 4 hours. The recombinant His-tagged protein were purified by nickel affinity chromatography according to the manufacturer's protocol (Bio-Rad) and the samples were stored at -70°C until use.

The Western Blot assay

To identify the expression of TK protein, the SDS-PAGE and Western Blotting assay were carried out according to the standard procedure. Antibodies against the DPV were raised in rabbits [50]. Briefly, the protein samples were electrophoresed in SDS-PAGE (12%) and subsequently transferred to PVDF membrane [51]. Then, Membrane was incubated with rabbit anti-DPV antibodies (1:200 dilution) overnight at 4°C. After washing three times with PBS-T (0.2% Tween 20 in PBS, PH 7.4), the second incubation of membrane was accomplished with horseradish peroxidase-linked goat anti-rabbit immunoglobulin G (IgG) (Amersham) for 1 hour at 37°C. As a result, the specific band was detected using DAB Enhancer Reagent Solution (Beijing Zhong Shan-Golden Bridge Biological Technology Co., Ltd., China).

Procedures for the TK-ELISA

The DPV-positive by inoculation of DPV Virulence Strain and the DPV-negative duck sera obtained from our laboratory were used to optimize the TK-ELISA. The 96-Well Microtiter Plates (Nunc, Denmark) were coated with 100 µl dilution of purified TK protein in sodium bicarbonate

buffer (pH 9.6), and were incubated at 4°C overnight. The optimal antigen concentration and the optimal serum dilution were determined by checkerboard titration of a coating antigen sample with gradient dilution (1:50, 1:100, 1:200, 1:400, 1:800, 1:1600) and an antisera sample with gradient dilution (1:20, 1:40, 1:80, 1:160). Plates were washed three times with PBS containing 0.05% Tween-20 (PBST) and then incubated with 100 µl of blocking solution (1% BSA in PBST) for 60 min at 37°C. Before adding the 100 µl of positive and negative diluted sera respectively, the plates, again, were washed and then incubated for 60 min at 37°C. After washing with PBST, 100 µl PBS diluted (1:1000, 1:2000, 1:4000), anti-duck horseradish peroxidase (KPL, Gaithersburg, USA) secondary antibody was added and the plates were incubated at 37°C for 45 min. Following the incubation, the plates were again washed as described above, and 100 µl/well of 3,3', 5,5'-etramethylbenzidine (TMB) was added. After 20 minutes incubation at 37°C, the reaction was stopped by addition of 50 µl of 2 mol/l H₂SO₄. The optical density (OD) was measured at 450 nm, using a Bio-Rad Model 860 Plate Reader (Bio-Rad, CA, USA).

The endpoint cut-off value

The 30 duck-negative sera samples were used to establish the endpoint cut-off value. The ELISA plates were coated with optimal TK antigen concentration and these sera samples at the optimal dilution were added. The TK-ELISA was performed in the optimal procedure described above. At the same time, the 0.01 M PBS was used for the blank control. Intra-day and inter-day variability were estimated by assaying sera samples containing 30 different duck-negative sera samples 6 times on the same day and on 3 separate days [52]. The data of result was analyzed with General Linear Model (GLM) procedures of SAS8.22, and the value was presented as least squares means± standard deviation. The significant differences of least square means were tested with the Duncan Test (*P* < 0.01). The cut-off value was calculated using the formula: mean of the negative serum values plus three standard deviations (SDs) [32].

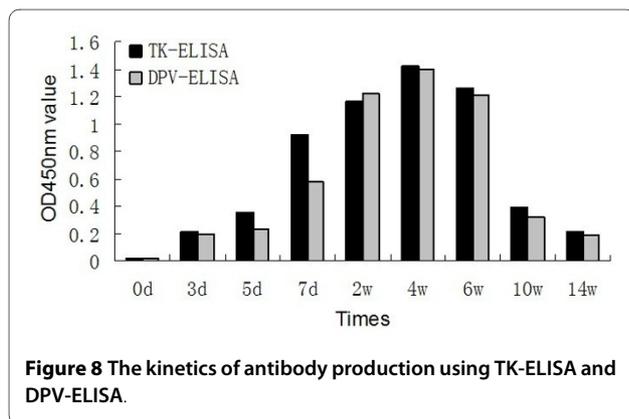


Figure 8 The kinetics of antibody production using TK-ELISA and DPV-ELISA.

Analytical specificity and sensitivity of the TK-ELISA

As stated above; the specificity of TK-ELISA was evaluated by testing with a panel of sera containing antibodies against DHBV, DHV, *R. A*, *E. coli*, and *S. anatum*. Other groups; the DPV-positive sera and DPV-negative sera were used as control groups respectively.

The 96-Well Microtiter Plates (Nunc, Denmark) were coated with 100 µl of a 1:200 (25 µg/100 µl) dilution of purified TK protein in sodium bicarbonate buffer (pH 9.6), and the seven diluted sera (1:160, 1:320, 1:640, 1:1280, 1:2560, 1:5120, 1:10,240) were added. The other operating conditions were carried out according to the optimal procedure described above. Therefore, the sensitivity of the TK-ELISA was inferred.

Analytical repeatability and reproducibility of TK-ELISA

Based on the validation of Serological Assays, the coefficient of variation (C.V.) for six replicates of 3 specimens were calculated for repeatability, these, according to TK-ELISA procedure were run on the same day. Also, the reproducibility of the set of specimens was analyzed in 3 runs during 3 days [53]. The data of result was analyzed with General Linear Model (GLM) procedures of SAS8.22, and the value was presented as least squares means ± standard deviation. The significant differences of least square means were tested with the Duncan Test.

Analytical inhibition of the TK-ELISA

Firstly, the DPV positive serum and the TK protein were diluted, respectively, with PBS, according to the optimal dilution (described above). Secondly, the dilution of serum was divided into two groups; A and B. Thirdly, the 50 µl dilution of TK protein was mixed with an equal volume of A (50 µl), and then the mixture was incubated for 60 min at 37°C. At last, the mixtures were tested in the TK-ELISA by adding 100 µl of each mixture, and the assay was performed according to the description above. At the same time, a 100 µl B was performed as the negative control and was also tested. The OD value was converted to percentage inhibition (PI) value using the formula [32]: $PI(\%) = 100 \times [1 - (OD_{\text{test serum}} / OD_{\text{control}})]$, where OD_{control} is the mean OD of wells containing positive serum alone.

The coincidence rate among SNT, TK-ELISA and DPV-ELISA

The suspected sera samples (n = 30) for infectious DPV were collected and preserved in our laboratory. These sera samples were tested by the TK-ELISA which is discussed above, SNT and the DPV-ELISA, respectively. The SNT was carried out for these samples in a Six-Well Cell Culture Plates with duck embryo fibroblast cells. Duplicates of serial twofold dilutions of sera, inactivated at 56°C for 30 min were tested as previously described

[54]. Titres were expressed as the reciprocal of the serum dilution that inhibited 75% of viral cytopathic effect. A serum sample was considered positive when it had a titer of = log₁₀1.0, equivalent to a serum dilution = 1:10. The DPV-ELISA was carried out for these samples referring to Xuefeng et al. [21]. Therefore, the coincidence rates were calculated from them.

Evaluation for the kinetics of antibody production using TK-ELISA and DPV-ELISA

Sera (n = 20) from twenty duck subcutaneously inoculated with attenuated live DPV vaccine (offered by author's laboratory), taken on days 0, 3, 5, 7 and weeks 2, 4, 6, 10, 14 after inoculation, were used by TK-ELISA and DPV-ELISA to study the kinetics of antibody production.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

YW carried out most of the experiments and wrote the manuscript. AC and MW critically revised the experiment design and the manuscript. HG, CS, SL, JX, RJ, DZ, XC, BL, HC and YZ helped with the experiment. All the authors read and approved the final manuscript.

Acknowledgements

The research were supported by grants from the Changjiang Scholars and Innovative Research Team in University (PCSIRT0848) and the earmarked fund for Modern Agro-industry Technology Research System (nycytx-45-12).

Author Details

¹Avian Diseases Research Center, College of Veterinary Medicine of Sichuan Agricultural University, Ya'an, Sichuan, 625014, China, ²Key Laboratory of Animal Diseases and Human Health of Sichuan Province, Ya'an, Sichuan, 625014, China and ³Epizootic Diseases Institute of Sichuan Agricultural University, Ya'an, Sichuan, 625014, China

Received: 9 February 2010 Accepted: 23 April 2010

Published: 23 April 2010

References

1. Baudet E: Een sterfte onder eenden in Nederland, veroorzaakt door een filterbaar virus (vogelpest). *Tijdschr Diergeneesk* 1923, **50**:455-459.
2. Fauquet CM, Mayo MA, Maniloff J, Desselberger U, Ball LA, (Eds): *Virus Taxonomy: Eighth Report of the International Committee on Taxonomy of Viruses*. California: Elsevier Academic Press; 2005.
3. Kaleta EF, Kuczka A, Kuhnhold A, Bunzenthall C, M BB, Hanka K, Redmann T, Yilmaz A: Outbreak of duck plague (duck herpesvirus enteritis) in numerous species of captive ducks and geese in temporal conjunction with enforced biosecurity (in-house keeping) due to the threat of avian influenza A virus of the subtype Asia H5N1. *German Veterinary Journal* 2007, **114**:3-11.
4. Weingarten M: Duck plague: clinical aspects, diagnosis, control. *Tierärztliche Praxis* 1989, **17**:53-56.
5. Leibovitz L, Hwang J: Duck plague on the American continent. *Avian Dis* 1968, **12**:361-378.
6. Converse KA, Kidd GA: Duck plague epizootics in the United States, 1967-1995. *Journal of wildlife diseases* 2001, **37**:347-357.
7. Campagnolo ER, Banerjee M, Panigrahy B, Jones RL: An outbreak of duck viral enteritis (duck plague) in domestic Muscovy ducks (*Cairina moschata domestica*) in Illinois. *Avian diseases* 2001, **45**:522-528.
8. Sluis Wvd: Ducks are a flavour for the future. *word poultry* 2004, **20**:41.
9. Huang YS, Au SC, Kong FJ, Lin WC: Investigations on Duck Plague Virus. *Journal of South China Agricultural College* 1980, **1**:21-36.

10. Burgess EC, Ossa J, Yuill TM: Duck plague: a carrier state in waterfowl. *Avian Dis* 1979, **23**:940-949.
11. Marius-Jestin V, Cherbonnel M, Picault JP, Bennejean G: Isolation from ducks of a hypervirulent strain of duck plague virus and an avian type 6 paramyxovirus. *Comp Immunol Microbiol Infect Dis* 1987, **10**:173-186.
12. Yufei G, Anchun C, Mingshu W, Chanjuan S, Renyong J, Shun C, Na Z: Development of TaqMan[®] MGB fluorescent real-time PCR assay for the detection of anatid herpesvirus 1. *Virology Journal* 2009, **6**:71.
13. Ji J, Du L, Xie Q, Cao Y, Zuo K, Xue C, Ma J, Chen F, Bee Y: Rapid diagnosis of duck plagues virus infection by loop-mediated isothermal amplification. *Research in veterinary science* 2009, **87**:53-58.
14. Renyong J, Anchun C, Mingshu W, Xuefeng Q, Dekang Z, Han G, Qihui L, Fei L, Yufei G, Xiaoyue C: Development and evaluation of an antigen-capture ELISA for detection of the UL24 antigen of the duck enteritis virus, based on a polyclonal antibody against the UL24 expression protein. *Journal of Virological Methods* 2009, **161**:38-43.
15. Islam MR, Khan MA: An immunocytochemical study on the sequential tissue distribution of duck plague virus. *Avian Pathol* 1995, **24**:189-194.
16. Guiping Y, Anchun C, Mingshu W, Fei L, Xiaoying H, Yonghong L, Chao X: Electron Microscopic Studies of the Morphogenesis of Duck Enteritis Virus. *Avian Diseases* 2008, **49**:50-55.
17. Toro H, Kaleta EF: Absence of neutralising antibodies to duck plague virus in the commercial duck and goose populations in West Germany (1980-1985). *Avian Pathol* 1986, **15**:57-62.
18. Bhanot V, Balamurugan V, Bhanuprakash V, Venkatesan G, Sen A, Yadav V, Yogisharadhya R, Singh RK: Expression of P32 protein of goatpox virus in *Pichia pastoris* and its potential use as a diagnostic antigen in ELISA. *J Virol Methods* 2009, **162**:251-257.
19. Iwaki K, Hayashi Y: Method for estimating the precision and detection limit for ELISA. *Nippon yakurigaku zasshi* 2009, **134**:207-211.
20. Kummitha CM, Mayle KM, Christman MA, Deosarkar SP, Schwartz AL, McCall KD, Kohn LD, Malgor R, Goetz DJ: A sandwich ELISA for the detection of Wnt5a. *J Immunol Methods* 2010, **352**:38-44.
21. Xuefeng Q, Anchun C, Mingshu W, Xiaoyan Y, Renyong J, Xiaoyue C: Development of an indirect-ELISA kit for detection of antibodies against duck plague virus. *Veterinary Science in China* 2007, **37**:690-694.
22. Anchun C, Mingshu W, Ming W, Weiguang Z, Yufei G, Renyong J, Chao X, Guiping Y, Yichen L: Construction of duck enteritis virus gene libraries and discovery, cloning and identification of viral nucleocapsid protein gene. *Chinese High Technology Letters* 2006, **16**:948-953.
23. Han G, Chao X, chun CA, shu WM, yong JIR, kang ZD, hui LQ, yue CX: Cloning of duck plague virus TK gene and analysis of its molecular characteristics. *Chinese Veterinary Science* 2008, **38**:297-302.
24. Li H, Liu S, Kong X: Characterization of the genes encoding UL24, TK and gH proteins from duck enteritis virus (DEV): a proof for the classification of DEV. *Virus Genes* 2006, **33**:221-227.
25. Neff C, Sudler C, Hoop RK: Characterization of western European field isolates and vaccine strains of avian infectious laryngotracheitis virus by restriction fragment length polymorphism and sequence analysis. *Avian Dis* 2008, **52**:278-283.
26. Shimojima Y, Jang HK, Ono M, Maeda K, Tohya Y, Mikami T: Identification and DNA sequence analysis of the Marek's disease virus serotype 2 genes homologous to the thymidine kinase and UL24 genes of herpes simplex virus type 1. *Virus Genes* 1997, **14**:81-87.
27. TC Mettenleiter: Aujeszky's disease (pseudorabies) virus: the virus and molecular pathogenesis--state of the art. *Vet Res* 2000, **31**:99-115.
28. Han MG, Kweon CH, Mo IP, Kim SJ: Pathogenicity and vaccine efficacy of a thymidine kinase gene deleted infectious laryngotracheitis virus expressing the green fluorescent protein gene. *Archives of Virology* 2002, **147**:1017-1031.
29. Coleman HM, Lima Bd, Morton V, Stevenson PG: Murine Gammaherpesvirus 68 Lacking Thymidine Kinase Shows Severe Attenuation of Lytic Cycle Replication In Vivo but Still Establishes Latency. *Journal of Virology* 2003, **77**:2410-2417.
30. Deshpande SS, (Ed): *Enzyme immunoassays: from concept to product development*. 1st edition. New York: Kluwer Academic Publishers; 1996.
31. Frazier CL, Shope RE: Detection of antibodies to alphaviruses by enzyme-linked immunosorbent assay. *J Clin Microbiol* 1979, **10**:583-585.
32. Ko YJ, Jeoung HY, Lee HS, Chang BS, Hong SM, Heo EJ, Lee KN, Joo HD, Kim SM, Park JH, Kweon CH: A recombinant protein-based ELISA for detecting antibodies to foot-and-mouth disease virus serotype Asia 1. *Journal of Virological Methods* 2009, **159**:112-118.
33. Chu JQ, Hu XM, Kim MC, Park CS, Jun MH: Development and validation of a recombinant nucleocapsid protein-based ELISA for detection of the antibody to porcine reproductive and respiratory syndrome virus. *Journal of microbiology (Seoul, Korea)* 2009, **47**:582-588.
34. Paweska JT, Jansen van Vuren P, Swanepoel R: Validation of an indirect ELISA based on a recombinant nucleocapsid protein of Rift Valley fever virus for the detection of IgG antibody in humans. *J Virol Methods* 2007, **146**:119-124.
35. Sueur JM, Beaumont K, Cabioch T, Orfila J, Betsou F: Diagnostic value of an ELISA using a recombinant 54-kDa species-specific protein from *Chlamydia pneumoniae*. *Clin Microbiol Infect* 2006, **12**:470-477.
36. Escribano JM, Pastor MJ, Sanchez-Vizcaino JM: Antibodies to bovine serum albumin in swine sera: implications for false-positive reactions in the serodiagnosis of African swine fever. *American journal of veterinary research* 1989, **50**:1118-1122.
37. Gill MB, Wright DE, Smith CM, May JS, Stevenson PG: Murid Herpesvirus-4 lacking thymidine kinase reveals route-dependent requirements for host colonization. *J Gen Virol* 2009, **90**:1461-1470.
38. Zabierowski SE, DeLuca NA: Stabilized Binding of TBP to the TATA Box of Herpes Simplex Virus Type 1 Early (tk) and Late (gc) Promoters by TFIIIA and ICP4. *American Society for Microbiology* 2008, **82**:3546-3554.
39. Shiau AL, Liu CW, Wang SY, Tsai CY: A simple selection system for construction of recombinant gD-negative pseudorabies virus as a vaccine vector. *Vaccine* 2002, **20**:1186-1195.
40. Miller J, Whetstone C, Bello L, Lawrence W: Determination of ability of a thymidine kinase-negative deletion mutant of bovine herpesvirus 1 to cause abortion in cattle. *Am J Vet Res* 1991, **52**:1038-1043.
41. McElroy AK, Albarino CG, Nichol ST: Development of a RVFV ELISA that can distinguish infected from vaccinated animals. *Virology journal* 2009, **6**:125.
42. Idrissi Bougrine S, Fassi Fihri O, el Harrak M, Fassi Fehri MM: Use of the immunoenzyme test ELISA-NS3 to distinguish horses infected by African horsesickness virus from vaccinated horses. *Revue scientifique et technique (International Office of Epizootics)* 1999, **18**:618-626.
43. Kit S, Awaya Y, Otsuka H, Kit M: Blocking ELISA to distinguish pseudorabies virus-infected pigs from those vaccinated with a glycoprotein gIII deletion mutant. *J Vet Diagn Invest* 1990, **2**:14-23.
44. Gold L: Expression of heterologous proteins in *Escherichia coli*. *Methods in enzymology* 1990, **185**:11-14.
45. Chao YP, Law W, Chen PT, Hung WB: High production of heterologous proteins in *Escherichia coli* using the thermo-regulated T7 expression system. *Applied microbiology and biotechnology* 2002, **58**:446-453.
46. Kleine T, Bartsch S, Blaser J, Schnierer S, Triebel S, Valentin M, Gote T, Tschesche H: Preparation of active recombinant TIMP-1 from *Escherichia coli* inclusion bodies and complex formation with the recombinant catalytic domain of PMNL-collagenase. *Biochemistry* 1993, **32**:14125-14131.
47. Steinitz M, Baraz L: A rapid method for estimating the binding of ligands to ELISA microwells. *Journal of Immunological Methods* 2000, **238**:143-150.
48. Gelman IH, Silverstein S: Identification of immediate early genes from herpes simplex virus that transactivate the virus thymidine kinase gene. *Proc Natl Acad Sci* 1985, **82**:5265-5269.
49. Honess RW, Craxton MA, Williams L, Gompels UA: A comparative analysis of the sequence of the thymidine kinase gene of a gammaherpesvirus, herpesvirus saimiri. *J Gen Virol* 1989, **70**:3003-3013.
50. Chao X, Anchun C, Mingshu W, Ming W: Study and application on cellular localization of DPV antigens in infected ducks by indirect immunoperoxidase technique. *Chin J Vet Sci* 2007, **27**:640-644.
51. Towbin H, Staehelin T, Gordon J: Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proc Natl Acad Sci USA* 1979, **76**:4350-4354.
52. Mire-Sluis AR, Barrett YC, Devanarayan V, Koren E, Liu H, Maia M, Parish T, Scott G, Shankar G, Shores E, et al.: Recommendations for the design and optimization of immunoassays used in the detection of host antibodies against biotechnology products. *J Immunol Methods* 2004, **289**:1-16.

53. Lau WH, Leong WS, Ismail Z, Gam LH: **Qualification and application of an ELISA for the determination of Tamm Horsfall protein (THP) in human urine and its use for screening of kidney stone disease.** *International journal of biological sciences* 2008, **4**:215-222.
54. Alvarado JF, Dolz G, Herrero MV, McCluskey B, Salman M: **Comparison of the serum neutralization test and a competitive enzyme-linked immunosorbent assay for the detection of antibodies to vesicular stomatitis virus New Jersey and vesicular stomatitis virus Indiana.** *J Vet Diagn Invest* 2002, **14**:240-242.

doi: 10.1186/1743-422X-7-77

Cite this article as: Wen *et al.*, A Thymidine Kinase recombinant protein-based ELISA for detecting antibodies to Duck Plague Virus *Virology Journal* 2010, **7**:77

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

